

## Effect On Growth of *Desmodesmus sp* Under Different Incubation Conditions

Wan Muhammad Ikram<sup>1</sup>, Fridelina Sjahri<sup>1</sup>, Noor Fazreen Dzulkafli<sup>1</sup>, Shuhaila Yaacob<sup>1</sup>, Kazuhiro Komatsu<sup>2</sup> Emi Fazlina Hashim<sup>3</sup>

Department of Science & Biotechnology, Faculty of Engineering & Life Science, University of Selangor, Jalan Timur Tambahan, 45600, Bestari Jaya, Selangor<sup>1</sup>

National Institute of Environmental Studies, Tsukuba, Japan<sup>2</sup>

Soka University Graduate School of Engineering, Tokyo, Japan<sup>3</sup>

Email: ikramzamri1995@gmail.com

**Abstract**— The aim of the research was to find the best conditions for selected microalgae species, *Desmodesmus. sp* in term of growth productivity in different pH, temperature and intensity of light. *Desmodesmus. Sp* (TRG-D01) was isolated from Setiu Wetlands, Terengganu and cultured using Bold Basal Media (BBM). The species was incubated under different selected parameters for nine days using microplate reader technique through Optical Density (OD) measurement. From the experiment, *Desmodesmus. sp* showed variety of growth pattern. Incubation under different pH, light intensity and temperature showed specific growth rate ranging from 0.21 to 0.47 d<sup>-1</sup> (maximum OD 0.67–0.93), 0.33 to 0.43 d<sup>-1</sup> (maximum OD 1.12) and 0.12 to 0.42 d<sup>-1</sup> (maximum OD=1.18) respectively. *Desmodesmus. sp* was optimum in were observed when incubated under pH in between 7 to 9, 56 to 91  $\mu\text{mol m}^{-2}\text{s}^{-1}$  of light intensity and optimum temperature between 23°C to 27°C.

**Keywords:** *Desmodesmus sp.*; Light Intensity; Optical Density; pH; Specific Growth Rate; Temperature

### 1. Introduction

Single celled green algae are photosynthetic microorganism that have ability to produce biomass from solar energy, CO<sub>2</sub> and nutrients. Nowadays, microalgae become an asset to consumers and industries especially from food industries, aquaculture and pharmaceutical. These products become interest for researcher for deeper understanding due to high nutritional value and potential that can be used in biodiesel, bioremediation human and animal nutrition. Generally stated that most microalgae able to undergoes photosynthesis and cell division process in between 15 to 30°C but best growth conditions would be in between 20 to 25°C (Munoz and Guieysse, 2006). Increasing temperature below optimum temperature showed that every 10°C increment, photosynthesis, cell division and growth should be expected to be double until maximum temperature reached (Difusa *et al.*, 2015). Exceeding favorable temperature would make microalgae growth decreased because heat would stress the enzyme or modify the protein that responsible for photosynthesis processes (Gardner *et al.*, 2011)

Light also one of the major elements that responsible for microalgae growth. Microalgae used light as source of energy for their metabolic processes. High amount of light exposed also damaged and irreversible damage maybe occurred as excessive amount of photon absorption exceed the electron turnover, thus inhibited the photosynthesis process (Ren, 2014). Light source not only effect the photosynthesis activities but also influenced the metabolic

pathway and cell composition (He *et al.*, 2015). Light intensity always dependent on volume and cell density of culture itself. On other words, light intensity needed to be increased if the microalgae cultured in bigger volume and high concentrations so that light able to penetrate entirely through the culture. Another critical conditions that determine microalgae growth is the pH since it determines the solubility and availability of CO<sub>2</sub> and nutrients content that significant for metabolic processes and the suitable range of pH for growth also specific for each microalgae (Rios *et al.*, 2015). pH of culture steadily increased due to utilization of inorganic carbon by microalgae. Depending on the species, microalgae able to adapt variety of environment conditions including pH, temperature and nutrient availability. Hence, the productivity of microalgae really dependent on how microalgae been cultured especially abiotic factor to maximize the production and quality in term of biological and chemical to design efficient photobioreactor (Van Wagenen *et al.*, 2014). This experiment was conducted to determine the optimum culture conditions namely pH, light intensity and temperature maximum growth of *Desmodesmus sp.* This work should be the one of first step for monitoring the best growth conditions before application into biomass production.

## 2. Materials & Methods

### *Microalgae Samples*

Identified microalgae, *Desmodesmus. sp* (TRG-D01) was isolated from Setiu Wetlands and donated by University Malaysia Terengganu (UMT) in broth stock culture. Bold Basal Media (BBM) based on (Bischoff, 1983) was prepared and used in this experiment. Each macro and micronutrients were made into stock culture at given concentrations to 0.934 L of distilled water. Final volume of media would be 1L. The BBM media final concentrations consist of  $2.94 \times 10^{-3}$  M NaNO<sub>3</sub>,  $3.04 \times 10^{-4}$  M MgSO<sub>4</sub>·7H<sub>2</sub>O,  $4.28 \times 10^{-4}$  M NaCl,  $4.31 \times 10^{-4}$  M K<sub>2</sub>HPO<sub>4</sub>,  $1.29 \times 10^{-3}$  M KH<sub>2</sub>PO<sub>4</sub>,  $1.70 \times 10^{-4}$  M CaCl<sub>2</sub>·2H<sub>2</sub>O, f/2 trace elements solution,  $1.85 \times 10^{-4}$  H<sub>3</sub>BO<sub>3</sub>,  $1.71 \times 10^{-4}$  Na<sub>2</sub>EDTA·2H<sub>2</sub>O,  $1.79 \times 10^{-5}$  FeSO<sub>4</sub>·7H<sub>2</sub>O,  $5.53 \times 10^{-4}$  KOH, f/2 vitamins solution (B<sub>1</sub>, Biotin and Cyanocobalamin).

The environment set up for initially growth of *Desmodesmus sp* were set according to (Komatsu *et al.*, 2019) with slight modification. The inoculated culture was incubated under controlled environment using climate chamber, Memmert HPP110. The initial surrounding condition of culture media was adjusted to pH 7 and inside climate chamber was set at temperature 25°C, 40  $\mu\text{mol m}^{-2}\text{s}^{-1}$  of light intensity with photoperiod 12H:12H of light and dark cycle and the humidity was set to 0%rh. The initial optical density before cultivation experiment were kept in between 0.01 to 0.05.

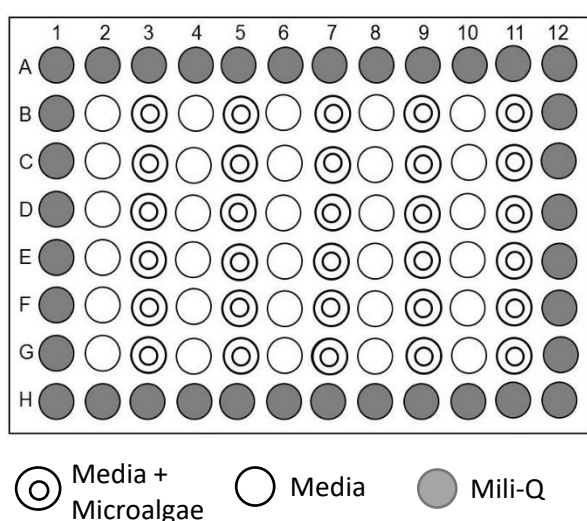
### *Manipulation of parameters*

For optimization purposes, the suitable range were selected based on different journal by (Difusa *et al.*, 2015; El-Sheekh *et al.*, 2017). For pH, the media was adjusted to pH 5, 6, 7, 8 and 9 by using either 1M of Hydrochloric Acid (HCl) or 1M of Sodium Hydroxide (NaOH). For light intensities and temperature, parameters were set according to preference by climate chamber. *Desmodesmus. sp* was cultivated under 5 different light intensities: (105  $\mu\text{mol m}^{-2}\text{s}^{-1}$ , 91  $\mu\text{mol m}^{-2}\text{s}^{-1}$ , 73  $\mu\text{mol m}^{-2}\text{s}^{-1}$ , 56  $\mu\text{mol m}^{-2}\text{s}^{-1}$  and 40  $\mu\text{mol m}^{-2}\text{s}^{-1}$ ). The optical length between light source to microplate was about 0.012 m and surrounded by white LED. The microplate arrangement was changed/adjusted every 24 hours after growth analysis to give the microplate equal irradiation. Cultivation under different temperature were set into 20, 23, 25

and 27°C. As the temperature increased, the microplate was wrapped with 2 layers of parafilm and Mili-Q water surrounding the well with algae need to be added daily. The precaution needed to be taken into note as the evaporation can happen as the temperature and light intensity increased.

### Microplate Technique

The incubation test started with inoculation of 20 µL of microalgae (10% of total volume per well) taken during exponential phases into 180 µL of cultured media. The total volume per well would be 200 µL. The analysis was conducted using 96 transparent with flat-bottom wells and microplate was divided into several zones as described in Figure 1. The border of microplate well was filled with 200 µL Mili-Q water to prevent evaporation. The border was not suitable to be used in experiment as it exposed to strong air flow though the microalgae able to grow due to more access to CO<sub>2</sub> and light. During 9 days of incubation, the pipetting (mixing) process all well containing microalgae have been conducted for efficient use of light and nutrients.



**Figure 1:** The designated zone of each wells using microplate reader for incubation experiment.

### Growth Analysis

Growth of *Desmodesmus. sp* was monitored through Optical Density (OD<sub>680</sub>) at 680 nm using Microplate reader Tecan M200 for every 24 hours in 9 days. Growth based of absorbance measurements were used in this study to determine microalgal biomass because the method was simple, fast, and proper technique to measure algal culture density (Sharma et al., 2016; Ding et al., 2015; Bohutskyi et al., 2015). For each recorded absorbance on each well, sensitive mean value (SMV) was applied using following formula:

$$SMV = \frac{OD_{tot} - OD_{Max} - OD_{Min}}{4}$$

where,

$OD_{tot}$  = Sum of absorbance at  $OD_{680}$  of 6 wells

$OD_{Max}$  = Maximum absorbance at  $OD_{680}$  of 6 wells

$OD_{Min}$  = Minimum absorbance  $OD_{680}$  of 6 wells

Average  $OD_{680}$  of wells containing microalgae were subtracted with well containing media only (control) to obtain exact  $OD_{680}$  of microalgae. Meanwhile, the specific growth rate was measured during exponential phases of growth. The specific growth rate ( $\mu$ ) and division rate ( $k$ ) was measured during exponential phases were calculated using formula as follow:

$$\mu = \frac{\ln(N_2/N_1)}{t_2 - t_1}$$

$$K = \frac{\mu}{\ln 2}$$

Where  $N_2$  and  $N_1$  are the value of OD at given times,  $t_2$  and  $t_1$ , respectively.

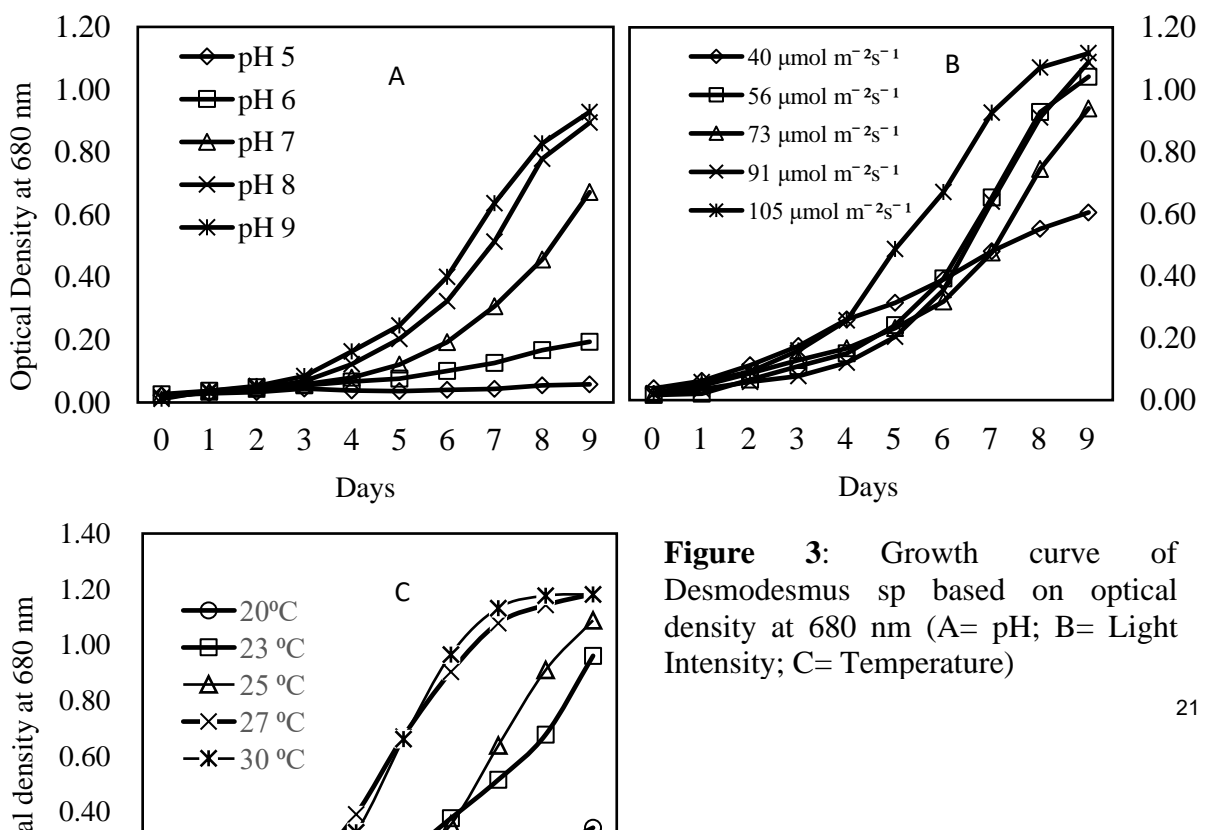
### 3. Results and Discussion

The maximum OD, specific growth rate and division rate were recorded and tabulated into Table 1 and growth curve of each different parameters were shown in Figure 3. Figure 3A shows the growth curve based on different pH tested on *Desmodesmus sp.* Among the studies of pH range of cultured media, it was observed that the microalgae favour the neutral to alkaline state, pH 7 to 9. The microalgae Within this range, optical density recorded ranging from 0.67 to 0.93 with specific growth rate of 0.39 to 0.44  $d^{-1}$  with division rate of 0.57 to 0.63. Within incubation period, no exponential phases or growth occurred when microalgae cultured under pH 5 and 6 of cultured media. Difusa *et al* (2015) reported that both isolated *Scenedesmus sp.* showed optimum growth at pH 7 to 9 in term of biomass productivity and lipid yield. Another study by (Bakuei *et al.*, 2015) also suggest that optimum growth for *Scenedesmus sp* ranged between 8.2 to 8.7. The nutrient uptake was rapidly efficient and released as ammonia during this condition most likely to be the reason for the significant increase in the alkaline state for growth in the presence of  $CO_2$ .

Based on Figure 3B, all levels of light intensity showed significant growth of *Desmodesmus* species. Result of study revealed that with maximum light intensity observed the exponential phases at day 4 with maximum  $OD_{680}$  at day 9 was 1.12 and specific growth rate of 0.43  $d^{-1}$ . In current study, *Desmodesmus sp.* seems able to grow varies of light intensity from 56-105  $\mu mol\ m^{-2}s^{-1}$  and lowering the light intensity directly slowed the growth of algae. This is due to the culture become denser and cell concentrations concentration become higher thus requires increasing light to penetrate throughout the culture depth (Wahidin *et al.*, 2013). as the microalgae grow and replicate, light shading must be taken into consideration as cell tend to block each other (Maynardo *et al.*, 2015). Incubation using microplate compared with flask or photobioreactor may be differ to one another as the volume and space of culture

exposed differently. Incubation under different light intensity in relation with growth rate can be applied in repeated batch cultures and relate the output to those large culture vessels and suitable for screening purposes (Van Wageningen *et al.*, 2014). Based on (Difusa *et al.*, 2015), both *Scenedesmus* species exhibit maximum biomass productivity when uniform increased of light intensity from  $27 \mu\text{mol m}^{-2}\text{s}^{-1}$  to  $81 \mu\text{mol m}^{-2}\text{s}^{-1}$  while increasing light intensity beyond  $81 \mu\text{mol m}^{-2}\text{s}^{-1}$  caused the biomass productivity became limited at  $91 \mu\text{mol m}^{-2}\text{s}^{-1}$ . However, the author found out that the lipid content achieved highest as light intensity increased.

The growth curve under different temperature showed in Figure 3C. At temperature at  $30^\circ\text{C}$  notified highest  $\text{OD}_{680}$ , 1.18 with specific growth rate of  $0.54 \text{ d}^{-1}$  and the exponential phase could be observed at day 4. Compared to other temperature points, the specific growth rates were ranging from 0.12 to  $0.44 \text{ d}^{-1}$  and the  $\text{OD}_{680}$  measured in between 0.34 to 1.18. Subsequently, the exponential phases of these temperature except for  $27^\circ\text{C}$  could be seen towards day 5 to day 6. However, at  $20^\circ\text{C}$ , the growth slowly increased at day 7 and maximum  $\text{OD}_{680}$  was 0.34 only till day 9. *Scenedesmus acutus* showed maximum growth and biomass productivity ( $0.42 \text{ g L}^{-1} \text{ d}^{-1}$ )  $30^\circ\text{C}$  Another study also reported that highest biomass productivity between temperature  $25^\circ\text{C}$  to  $30^\circ\text{C}$  and in between  $25^\circ\text{C}$  to  $30^\circ\text{C}$  with no significant variation but the growth inhibit when cultivated over  $30^\circ\text{C}$  (El-Sheekh *et al.*, 2017). Higher temperature which is favour for microalgae in current study able to supply thermal energy that influenced cellular enzymatic activities, nutrient uptake and  $\text{CO}_2$  fixation cycle (Karlo *et al.*, 2015). Vanags *et al* (2015) reported that optimum temperature in between  $23^\circ\text{C}$  to  $27^\circ\text{C}$  ( $25^\circ\text{C} \pm$ ) for *Desmodesmus communis* in shake flask culture with no significant difference between the range. The author also stated that  $21^\circ\text{C}$  and  $29^\circ\text{C}$  inhibited the growth thus not suitable for cultivation conditions. The contradiction of result maybe resulted from the cultivation technique used in study.



**Figure 3:** Growth curve of *Desmodesmus* sp based on optical density at 680 nm (A= pH; B= Light Intensity; C= Temperature)

**Table 1:** The maximum OD, specific growth rate and division rate of *Desmodesmus sp* at different incubation conditions

Parameters		Maximum OD at 680 nm	Specific Growth Rate, $d^{-1}$	Division rate, $k$
pH	5	$0.06 \pm 0.01$	NA	NA
	6	$0.19 \pm 0.03$	NA	NA
	7	$0.67 \pm 0.04$	$0.39 \pm 0.02$	$0.57 \pm 0.03$
	8	$0.89 \pm 0.02$	$0.44 \pm 0.02$	$0.63 \pm 0.03$
	9	$0.93 \pm 0.02$	$0.40 \pm 0.02$	$0.58 \pm 0.02$
Light Intensity ( $\mu\text{mol m}^{-2}\text{s}^{-1}$ )	40	$0.60 \pm 0.08$	$0.18 \pm 0.07$	$0.26 \pm 0.10$

	56	$1.04 \pm 0.04$	$0.43 \pm 0.02$	$0.62 \pm 0.03$
	73	$0.94 \pm 0.05$	$0.34 \pm 0.01$	$0.50 \pm 0.02$
	91	$1.09 \pm 0.04$	$0.46 \pm 0.09$	$0.67 \pm 0.12$
	105	$1.12 \pm 0.02$	$0.43 \pm 0.02$	$0.62 \pm 0.02$
Temperature (°C)	20	$0.34 \pm 0.08$	$0.25 \pm 0.07$	$0.36 \pm 0.11$
	23	$0.96 \pm 0.07$	$0.30 \pm 0.05$	$0.43 \pm 0.07$
	25	$1.09 \pm 0.04$	$0.44 \pm 0.09$	$0.63 \pm 0.12$
	27	$1.18 \pm 0.05$	$0.42 \pm 0.02$	$0.42 \pm 0.03$
	30	$1.18 \pm 0.04$	$0.54 \pm 0.02$	$0.78 \pm 0.04$

NA=Not Available; no exponential phases of growth observed within incubation period  
Note: The value represent means  $\pm$  SD (n=6)



#### 4. Conclusions

*Desmodesmus sp* (TRG-D01) able to grow in different conditions, at least in the experiment conducted. Present experiment clearly observed that the species favours in neutral to alkaline state (pH 7, 8 and 9) of cultured media, irradiance levels in between  $56 \mu\text{mol m}^{-2}\text{s}^{-1}$  to  $105 \mu\text{mol m}^{-2}\text{s}^{-1}$ , and the temperature ranging from  $23^{\circ}\text{C}$  to  $27^{\circ}\text{C}$  give the steady increased of growth in term of optical density. Since the current study only cover the growth aspect and limited study due to incubation technique, it could be useful in future to investigate biochemical composition such as lipid content based on microplate technique. This groundwork could be useful as a first step towards biomass production.

#### ACKNOWLEDGEMENT

This research was supported by Japan Science and Technology Agency (JST)/Japan International Cooperation Agency (JICA), Science and Technology Research Partnership for Sustainable Development (SATREPS) through the project for Continuous Operation System for Microalgae Production Optimized for Sustainable Tropical Aquaculture (COSMOS; Grant No. [JPMJSA1509](#)), and the SATREPS-COSMOS Matching Fund from the Ministry of Higher Education, Malaysia (MOHE).

#### 5. References

- Bakuei, N., Amini, G., Najafpour, G. D., Jahanshahi, M., & Mohammadi, M. (2015). Optimal cultivation of *Scenedesmus sp.* microalgae in a bubble column photobioreactor.
- Bohutskyi, P., Liu, K., Nasr, L. K., Byers, N., Rosenberg, J. N., Oyler, G. A., ... & Bouwer, E. J. (2015). Bioprospecting of microalgae for integrated biomass production and phytoremediation of unsterilized wastewater and anaerobic digestion centrate. *Applied microbiology and biotechnology*, 99(14), 6139-6154.
- Demirel, Z., Yilmaz, F. F., Ozdemir, G., & Dalay, M. C. (2018). Influence of Media and Temperature on the Growth and the Biological Activities of *Desmodesmus protuberans* (FE Fritsch & MF Rich) E. Hegewald. *Turkish Journal of Fisheries and Aquatic Sciences*, 18(10), 1195-1203.
- Difusa, A., Talukdar, J., Kalita, M. C., Mohanty, K., & Goud, V. V. (2015). Effect of light intensity and pH condition on the growth, biomass and lipid content of microalgae *Scenedesmus* species. *Biofuels*, 6(1-2), 37-44.
- Ding, G. T., Takriff, M. S., Salihon, S., Syukri, M., & Rahaman, A. B. D. (2015). Feasibility of the optical density (OD) in the determination of the microalgal biomass using palm oil mill effluent (POME) as medium. In *Proceedings of 50th the IIER International Conference, Zurich, Switzerland, 26th December*.
- El-Sheekh, M., Abomohra, A. E. F., Abd El-Azim, M., & Abou-Shanab, R. (2017). Effect of temperature on growth and fatty acids profile of the biodiesel producing microalga *Scenedesmus acutus*. *BASE*.



- He, Q., Yang, H., Wu, L., & Hu, C. (2015). Effect of light intensity on physiological changes, carbon allocation and neutral lipid accumulation in oleaginous microalgae. *Bioresource technology*, 191, 219-228.
- Komatsu, Kazuhiro., Onouchi, Hidemi., Imai, Akio., Kawasaki, Nobuyuki., Hashim, Emi & Rajuddin, Mohd. (2019). Effects of Dissolved Organic Matter in Soil Extracts on the Growth of Microalgae 土壤抽出物中溶存有機物 (DOM) の微細藻類増殖に及ぼす影響. *Journal of Japan Society on Water Environment*. 42. 239-246. 10.2965/jswe.42.239.
- Maynardo, J. J., Doshi, V. E. E. N. A., Rajanren, J. R., & Rajasekaran, R. (2015). The Optimization of Light Intensity and Drying Temperature on Lipid Content of Microalgae *Nannochloropsis oculata*. *J Eng Sci Technol*, 112-121.
- Munoz, R., & Guieysse, B. (2006). Algal–bacterial processes for the treatment of hazardous contaminants: a review. *Water research*, 40(15), 2799-2815.
- Ren, T. (2014). Primary factors affecting growth of microalgae optimal light exposure duration and frequency. Master thesis, Iowa State University.
- Rios, L. F., Klein, B. C., Luz, L. F., Maciel Filho, R., & Maciel, M. W. (2015). Nitrogen starvation for lipid accumulation in the microalga species *Desmodesmus* spp. *Applied biochemistry and biotechnology*, 175(1), 469-476. <https://dx.doi.org/10.1007/s12010-014-1283-6>
- Sharma, A. K., Sahoo, P. K., Singhal, S., & Patel, A. (2016). Impact of various media and organic carbon sources on biofuel production potential from *Chlorella* spp. *3 Biotech*, 6(2), 116.
- Van Wagenen, J., Holdt, S. L., De Francisci, D., Valverde-Pérez, B., Plósz, B. G., & Angelidaki, I. (2014). Microplate-based method for high-throughput screening of microalgae growth potential. *Bioresource technology*, 169, 566-572.
- Vanags, J., Kunga, L., Dubencovs, K., Galvanauskas, V., & Grīgs, O. (2015). Influence of light intensity and temperature on cultivation of microalgae *Desmodesmus Communis* in flasks and laboratory-scale stirred tank photobioreactor. *Latvian Journal of Physics and Technical Sciences*, 52(2), 59-70.
- Wahidin, S., Idris, A., & Shaleh, S. R. M. (2013). The influence of light intensity and photoperiod on the growth and lipid content of microalgae *Nannochloropsis* sp. *Bioresource technology*, 129, 7-11.